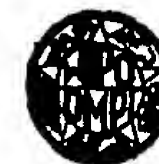


**PCT**WORLD INTELLECTUAL PROPERTY ORGANIZATION  
International Bureau**EA**

## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification <sup>6</sup> :</b> <b>G01N 33/543 // G01B 7/34</b>	<b>A1</b>	<b>(11) International Publication Number:</b> <b>WO 96/31775</b> <b>(43) International Publication Date:</b> 10 October 1996 (10.10.96)
<b>(21) International Application Number:</b> PCT/SE96/00431 <b>(22) International Filing Date:</b> 2 April 1996 (02.04.96)  <b>(30) Priority Data:</b> 9501236-5      4 April 1995 (04.04.95)      SE  <b>(71)(72) Applicants and Inventors:</b> OSCARSSON, Sven [SE/SE]; Dalgatan 7B, S-752 28 Uppsala (SE). BERGMAN, Anna [SE/SE]; Slottsgränd 8A, S-753 09 Uppsala (SE). QUIST, Arjan [NL/SE]; Rackarbergsgatan 34:155, S-752 32 Uppsala (SE). REIMANN, Curt, T. [US/SE]; Bredgränd 17, S-753 20 Uppsala (SE). SUNDQVIST, Bo [SE/SE]; Ihres väg 17, S-752 63 Uppsala (SE).  <b>(74) Agents:</b> WIDEGREN, Ann-Charlotte et al.; Bergensträhle & Lindvall AB, P.O. Box 17704, S-118 93 Stockholm (SE).		<b>(81) Designated States:</b> CA, JP, US, European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).  <b>Published</b> <i>With international search report.</i>
<b>(54) Title:</b> A METHOD FOR DETECTING AND QUANTIFYING ANALYTES BY MEANS OF SCANNING FORCE MICROSCOPY  <b>(57) Abstract</b>  Analytes are detected and quantified without the use of labels. A solution containing a substance capable of forming an immunocomplex is brought into contact with a support surface on which the other component of the immunocomplex to be formed is immobilized and the formed complex is detected and distinguished from substances which do not form part of the immunocomplex by means of scanning force microscopy.		

**FOR THE PURPOSES OF INFORMATION ONLY**

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AM	Armenia	GB	United Kingdom	MW	Malawi
AT	Austria	GE	Georgia	MX	Mexico
AU	Australia	GN	Guinea	NE	Niger
BB	Barbados	GR	Greece	NL	Netherlands
BE	Belgium	HU	Hungary	NO	Norway
BF	Burkina Faso	IE	Ireland	NZ	New Zealand
BG	Bulgaria	IT	Italy	PL	Poland
BJ	Benin	JP	Japan	PT	Portugal
BR	Brazil	KE	Kenya	RO	Romania
BY	Belarus	KG	Kyrgyzstan	RU	Russian Federation
CA	Canada	KP	Democratic People's Republic of Korea	SD	Sudan
CF	Central African Republic	KR	Republic of Korea	SE	Sweden
CG	Congo	KZ	Kazakhstan	SG	Singapore
CH	Switzerland	LI	Liechtenstein	SI	Slovenia
CI	Côte d'Ivoire	LK	Sri Lanka	SK	Slovakia
CM	Cameroon	LR	Liberia	SN	Senegal
CN	China	LT	Lithuania	SZ	Swaziland
CS	Czechoslovakia	LU	Luxembourg	TD	Chad
CZ	Czech Republic	LV	Latvia	TG	Togo
DE	Germany	MC	Monaco	TJ	Tajikistan
DK	Denmark	MD	Republic of Moldova	TT	Trinidad and Tobago
EE	Estonia	MG	Madagascar	UA	Ukraine
ES	Spain	ML	Mali	UG	Uganda
FI	Finland	MN	Mongolia	US	United States of America
FR	France	MR	Mauritania	UZ	Uzbekistan
GA	Gabon			VN	Viet Nam

A method for detecting and quantifying analytes by means of  
scanning force microscopy

The present invention relates to a method for detecting and optionally quantifying analytes which are capable of forming immunocomplexes. More particularly the invention relates to such a method wherein no labelling of substances is used and wherein the immunocomplexes are formed on a support surface and then detected and optionally quantified by means of scanning force microscopy.

The most sensitive analytical techniques available today for detecting immunoassay complexes are radioimmunoassay (RIA) and enzyme-linked immunosorbent assay (ELISA). Both of these techniques are used for example for testing for drug abuse. The RIA-test is very sensitive but has the disadvantage that a radioactive labelling must be used. In ELISA testing procedures detection is made by a simple colorimetric assay. The ELISA procedure is, however, usually less sensitive than the RIA procedure.

Scanning force microscopy (SFM) for three-dimensional imaging of macromolecules was introduced in 1986 and has since then found increasing interest and use in the investigation of biomolecules. The two modes of SFM, the contact mode (CM) and the more gentle tapping mode (TM), have for example been used for investigation of the coiling of DNA, the structure of human serum albumin etc, adsorbed on support surfaces.

According to the present invention it has been found that it is possible to detect and optionally quantify analyte molecules which are capable of forming immunocomplexes by forming the complexes on a support surface and then detecting and quantifying the formed immunocomplexes by means of SFM.

As mentioned above the RIA-procedure requires labelling with radioactive substances and in other biotechnical processes labelling of substances with e.g. gold or fluorescent compounds for detection is often used. The present invention is especially advantageous in that it provides an ultrasensitive technique for detecting immunocomplexes in which no labelling of substances is required. Extremely small amounts of substances can be detected, and even single analyte mole-

cules.

The invention thus relates to a method as defined in the appended claims.

The present method is based on the finding that it is possible to detect and quantify analytes which are capable of forming immunocomplexes using SFM but without use of any special labelling for the detection.

The term "analyte" as used herein is intended to encompass the antigen or the hapten which is to be detected. The term "immunocomplex" as used herein is intended to encompass complexes formed by an antigen and an antibody or by a hapten and an antibody. Haptens are low molecular weight substances which can react with the effector cells of the immune response (humoral antibodies or stimulated T-lymphocytes). As some examples of haptens can be mentioned drugs and drug metabolites, hormones etc.

In the present method the immunocomplexes are formed by bringing a solution containing a substance capable of forming an immunocomplex into contact with a support surface on which the other component of the immunocomplex to be formed is immobilized. Hereby either analytes or antibodies against an analyte can be immobilized on the surface. In the first case, when analytes are immobilized, the solution which is brought into contact with the support surface thus contains antibodies. Usually, and preferably, antibodies will be immobilized on the support surface and a solution containing the analyte brought into contact with this. The solution can for example be a body fluid which is to be analyzed for the presence of medicaments or drug abuse. Other instances wherein the present method can be used are for example in analysis of waste fluids, for example from industrial processes, which are to be investigated with regard to toxic substances such as phenol, styrene, acrylic acid, thiazoles etc.

One way of quantifying the analytes is to let the surface with the immobilized antibody directed to the analyte gradually pass through the sample. Depending on the amount of analyte in the liquid sample immunocomplexes will be formed over varying areas of the support surface or along different distances on this, which can be used for quantitative diag-

nosis of the analyte.

The surface on which the detection is made is preferably organised in such a manner that it can accommodate a maximum number of immunocomplexes and this can for example be carried out by means of a multipoint applicator on a nanometer scale or by masking the surface to allow maximum number of complexes to be adsorbed on or bound to the surface.

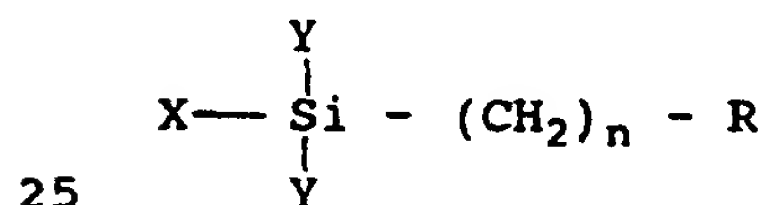
When a sample is to be analyzed the solution containing the analyte is brought into contact with the surface, on which the other component of the immunocomplex to be formed is immobilised, in such a manner as to wet the surface. It might often be necessary to use very highly diluted samples. The actual operation of the scanning force microscopy in tapping mode can be carried out in air or a liquid cell can be used. In the latter operation manner the tip of the instrument is inserted in the liquid cell which is then placed on the sample with a sealing O-ring between the cell and the sample substrate. As in the tapping mode in air the sample is mounted on top of the scanner. Since the fluid medium tends to damp the cantilevers normal resonant frequency, the entire liquid cell can be oscillated to drive the cantilever into oscillation. Further operation is similar to the operation of the tapping mode in air, which is well known to the man skilled in the art. The liquid cell is equipped with an inlet- and outlet tube which makes it possible to inject different solutions while operating the microscope.

For SFM-techniques it is important that the solid support surface is extremely smooth and plane to prevent the surface from interfering with the complexes to be analyzed. The support surface can be organic or inorganic. As examples of organic support surfaces can be mentioned teflon® and polystyrene. Usually inorganic supports are used. A support surface of glass can be used but usually smoother flat support surfaces of mica, graphite, gypsum, polished silicon, silicon wafers, or various forms of crystals which form an atomically flat surface are employed. Mica is a very suitable support surface. It has a surface roughness of  $\approx 1\text{\AA}$  (Ångström) and a new clean surface can be exposed simply by peeling off a few layers of mica for example with Scotch tape.

Generally speaking the surfaces used as supports in SFM-techniques have to be specially treated, for example by extremely careful washing, rinsing, drying etc, and often by chemical modification, in order that the desired imaging will be  
 5 obtained on a level that allows separation of molecules, aggregates, complexes etc.

It is possible to utilize the present method with support surfaces as above without derivatization, i.e. chemical modification, whereby haptens, antigens or antibodies will be  
 10 immobilized on the surface through adsorption. It is, however, preferred that the support surfaces are derivatized so that a covalent bonding is obtained and thus a higher reliability in the detection and quantifying. It is likewise preferred to use the tapping mode SFM for a higher reliability  
 15 since there is a risk that the contact mode SFM will push away compounds and complexes, particularly of higher molecular weight, from the support surface.

The support surfaces can be derivatized by per se known reagents as used for example for inorganic support materials  
 20 for chromatography. A common method is silanization. The general structure of the silane reagent for covalent immobilization of proteins on inorganic surfaces is



wherein

X and Y = Cl or X = Cl and Y = H; or

X and Y = alkyl or alkoxy groups with 1 or 2 carbon atoms;

n = 1 - 8, preferably 3 and

30 R is a functional group to which the naturally reactive groups of proteins can bind, such as amino or thiol groups of the protein or thiol groups introduced into the protein. Suitable functional groups R are -CH=CH<sub>2</sub>, an epoxy group, NH<sub>2</sub>, SH, pyridine or S-pyridine. Commercially available si-  
 35 lanes can usually be chemically modified to change the functional group R to the in each case appropriate functional group, if required. Several protocols for silanization are known from the literature.

The reagent for derivatization of the surface can also be

a hydrocarbon compound corresponding to the above given formula, ie of the same formula but with a carbon atom instead of the silicon atom. Another way of modifying the support surface is by covalent binding of organic polymers to the surface. Thus examples of immobilization of proteins via polymers such as polyethylene glycol and dextran are known from the literature. These known methods must usually be optimized and modified for use with SFM.

As mentioned above several protocols for silanization are known and also for other types of derivatization. Derivatization of mica can for example be carried out in liquid or in vapours. In one liquid process a piece of mica is peeled on both sides and put in a freshly prepared solution of the derivatizing reagent in a suitable solvent, such as for example toluene. The modified mica is then rinsed with the same solvent and dried, for example using a flow of nitrogen. Methods for derivatization in vapours for different reagents have been described. A piece of mica can for example be peeled on both sides and placed in a desiccator which also contains a small amount of the reagent and the desiccator is then placed under vacuum for a certain time.

#### Example 1

In this experiment the formation of biocomplexes, or the lack thereof, between single human serum albumin molecules (HSA) and single or multiple antibodies, rabbit anti-human serum albumin (a-HSA), and human immunoglobulin G (IgG) adsorbed on mica surfaces was investigated. SFM probing and studies were made with TM-SFM (Nanoscope III®, Digital Instruments Inc., Santa Barbara, CA, USA) using tips with an end-radius of about 10 nm.

HSA, a-HSA and IgG were dissolved in tris buffer (pH 7.4) at concentrations of 2.5 µg/ml, 13.4 µg/ml and 0.7 µg/ml respectively. The concentrations were selected to give roughly the same area density of molecules (40-60 per µm<sup>2</sup>) adsorbed on the surface. In each given exposure a 50 µl volume of the protein solution was placed on a freshly-cleaved mica surface (Muscovite green mica from Asheville-Schoonmaker Mica Co. Newport News VA, USA). The solution was spread out over approximately 1 cm<sup>2</sup>, allowed to remain on the surface for 5 min-



utes and subsequently rinsed away with 1 ml of tris buffer. The surface was then dried using a flow of nitrogen and probed with TM-SFM.

To study the antigen-antibody interaction the mica surfaces were first exposed to one protein solution, rinsed with 1 ml tris buffer, dried and studied by TM-SFM. The surfaces were then exposed to the second protein solution (without recleaving), rinsed, dried and studied again by TM-SFM. Each TM-SFM image covered an area of  $0.5 \mu\text{m} \times 0.5 \mu\text{m}$ . The height histograms drawn on basis of the different TM-SFM images showed large differences which made it possible to quantitatively distinguish between different species on the surfaces. This is shown in the Table below which gives peak positions and full widths at half maximum (FWHM).

<u>Table</u>		
<u>Experiment</u>	<u>1st Peak pos. ± FWHM (nm)</u>	<u>2nd Peak pos. ± FWHM (nm)</u>
A HSA	$0.62 \pm 0.28$	
B a-HSA	$1.91 \pm 0.66$	
C IgG	$1.75 \pm 0.60$	
D HSA exposed to IgG	$\approx 0.62$	$\approx 1.75$
E HSA exposed to a-HSA	$\approx 0.62$	$3.03 \pm 1.60$
F IgG exposed to HSA	$\approx 0.62$	$1.35 \pm 0.72$
G a-HSA exposed to HSA	$\approx 0.62$	$1.38 \pm 0.80$

\* Note: In experiments E, F and G the HSA contribution has been subtracted for the 2nd peak.

The following remarks are given to further clarify the results given in the Table above.

Although the height information obtained by TM-SFM is not totally topological, the height histogram showed large differences which allowed quantitative distinction between the presence of different species on the surfaces. In experiment D, wherein HSA preadsorbed on the mica surface was exposed to IgG two distinct populations of molecules, within height ranges corresponding to those of separately adsorbed HSA and IgG were observed, and no interaction between HSA and human IgG could be observed. In experiment E, wherein HSA on mica



was exposed to a-HSA, larger features, both with regard to lateral dimensions and height, were observed than with either HSA or a-HSA absorbed separately on mica. The image showed the expected complexing, since the a-HSA is raised in rabbit  
5 to specifically interact with HSA. In the results shown for experiment E the HSA contribution for the 2nd peak has been subtracted.

The experiments further showed that HSA binds spontaneously and irreversibly to hydrophilic mica surfaces which  
10 were not derivatized, even though the net charge of HSA as well as the surface charge are negative. IgG did, however, bind more weakly to the mica and would thus preferably be immobilized on a derivatized surface.

Claims

1. A method for label-free detecting and optionally quantifying of analytes, characterized in that a solution containing a substance capable of forming an immunocomplex is brought into contact with a support surface on which the other component of the immunocomplex to be formed is immobilized and that the formed complex is detected and distinguished from substances which do not form part of the immunocomplex by means of scanning force microscopy.

2. A method according to claim 1, characterized in that the two components forming the immunocomplex are antibodies and antigens.

3. A method according to claim 1 or 2, characterized in that the solution contains the substance to be detected and that this an antigen.

4. A method according to claim 1, characterized in that the solution contains the substance to be detected and that this a low molecular drug or a hormone.

5. A method according to any of the preceding claims, characterized in that the support surface is of glass, mica or polished silicon.

6. A method according to claim 5, characterized in that the support surface is chemically modified for immobilization by covalent binding.

7. A method according to claim 6, characterized in that the support surface is silanized.

8. A method according to any of the preceding claims, characterized in that tapping mode scanning force microscopy is used for the detection.

9. A method according to any of the preceding claims, characterized in that the scanning force microscope is operated in air.

## INTERNATIONAL SEARCH REPORT

International application No.

PCT/SE 96/00431

## A. CLASSIFICATION OF SUBJECT MATTER

IPC6: G01N 33/543 // G01B 7/34

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC6: G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

SE,DK,FI,NO classes as above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EPODOC, WPI, US PATFULL

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	Nucleic Acids Research, Volume 22, No 16, 1994, Lia I. Pietrasanta et al, "Probing specific molecular conformations with the scanning force microscope. Complexes of plasmid DNA and anti-Z-DNA antibodies", page 3288 - page 3292, See "materials and methods", page 3289  --	1
X	Proc.Natl.Acad.Sci., Volume 92, February 1995, (USA), Sándor Damjanovich et al, "Structural hierarchy in the clustering of HLA class I molecules in the plasma membrane of human lymphoblastoid cells", page 1122 - page 1126, See "materials and methods", page 1122-1123  --	1



Further documents are listed in the continuation of Box C.



See patent family annex.

## \* Special categories of cited documents

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&amp;" document member of the same patent family

Date of the actual completion of the international search

5 July 1996

Date of mailing of the international search report

17.07.96

Name and mailing address of the ISA/

Swedish Patent Office

Box 5055, S-102 42 STOCKHOLM

Facsimile No. +46 8 666 02 86

Authorized officer

Carl-Olof Gustafsson

Telephone No. +46 8 782 25 00

## INTERNATIONAL SEARCH REPORT

International application No.

PCT/SE 96/00431

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	National Library of Medicine, file Medline, Medline accession no. 94146259, Karrasch S. et al: "Covalent binding of biological samples to solid supports for scanning probe microscopy in buffer solution", Biophys J 1993 Dec;65(6): 2437-46  --	1-7
A	Dialog Information Services, File 73, EMBASE, Dialog accession no. 9307864, EMBASE accession no. 94262429, Warkentin P. et al: "Differential surface binding of albumin, immunoglobulin G and fibrinogen", BIOMATERIALS (United Kingdom), 1994, 15/10 (786-795)  --	1-9
X	US 5412980 A (VIRGIL B. ELINGS ET AL), 9 May 1995 (09.05.95), column 1 - column 4; column 7, line 60 - column 8, line 23  --	1-9
X	US 5363697 A (TOHRU NAKAGAWA), 15 November 1994 (15.11.94), see example 2 and column 1-4  --	1-7
X	US 5372930 A (RICHARD J. COLTON ET AL), 13 December 1994 (13.12.94), see examples and claims  --	1-7
X	US 5322769 A (TIMOTHY J. BOLLING ET AL), 21 June 1994 (21.06.94), column 9 - column 10  --	1-7
A	US 5289004 A (TAKAO OKADA ET AL), 22 February 1994 (22.02.94), column 1 - column 2  --	1-9
A	US 5156810 A (HANS O. RIBI), 20 October 1992 (20.10.92), column 16, line 16 - line 29  -- -----	1

# INTERNATIONAL SEARCH REPORT

Information on patent family members

01/04/96

International application No.

PCT/SE 96/00431

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US-A- 5412980	09/05/95	EP-A- 0587459 JP-A- 7270434	16/03/94 20/10/95
US-A- 5363697	15/11/94	EP-A,A- 0511662 JP-A- 5093623 JP-A- 4330281	04/11/92 16/04/93 18/11/92
US-A- 5372930	13/12/94	NONE	
US-A- 5322769	21/06/94	AT-T- 131876 AU-A- 3120689 CA-A- 1335358 DE-D- 68925138 EP-A,A,A 0331961 SE-T3- 0331961 ES-T- 2083957 JP-A- 2035089 JP-B- 8013273 US-A- 5124255 US-A- 5312737 AU-A- 4643693 CA-A- 2136765 EP-A- 0647277 JP-T- 7508411 WO-A- 9400594	15/01/96 14/09/89 25/04/95 00/00/00 13/09/89  01/05/96 05/02/90 14/02/96 23/06/92 17/05/94 24/01/94 06/01/94 12/04/95 21/09/95 06/01/94
US-A- 5289004	22/02/94	EP-A,A,A 0449221 JP-A- 4212252 JP-A- 4270903 JP-A- 5141961	02/10/91 03/08/92 28/09/92 08/06/93
US-A- 5156810	20/10/92	CA-A- 2019039 EP-A,A,A 0402917 JP-A- 3128449 US-A- 5268305 US-A- 5427915 US-A- 5491097	15/12/90 19/12/90 31/05/91 07/12/93 27/06/95 13/02/96

